



Synthesis and characterisation of nanophase hydroxyapatite co-substituted with strontium and zinc

Lowry, N., Brolly, M., Han, Y., McKillop, S., Meenan, B., & Boyd, A. (2018). Synthesis and characterisation of nanophase hydroxyapatite co-substituted with strontium and zinc. *Ceramics International*, 44(7), 7761-7770. <https://doi.org/10.1016/j.ceramint.2018.01.206>

[Link to publication record in Ulster University Research Portal](#)

Published in:
Ceramics International

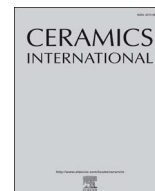
Publication Status:
Published (in print/issue): 31/05/2018

DOI:
[10.1016/j.ceramint.2018.01.206](https://doi.org/10.1016/j.ceramint.2018.01.206)

Document Version
Author Accepted version

General rights
Copyright for the publications made accessible via Ulster University's Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The Research Portal is Ulster University's institutional repository that provides access to Ulster's research outputs. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact pure-support@ulster.ac.uk.



Synthesis and characterisation of nanophase hydroxyapatite co-substituted with strontium and zinc

N. Lowry*, M. Brolly, Y. Han, S. McKillop, B.J. Meenan, A.R. Boyd

Nanotechnology and Integrated Bioengineering Centre (NIBEC), School of Engineering, Ulster University, Northern Ireland, UK

ARTICLE INFO

Keywords:

Bioceramic
Nano-hydroxyapatite
Co-substitution
Strontium
Zinc

ABSTRACT

In order to develop new bioactive calcium phosphate (CaP) materials to repair bone defects, it is important to ensure these materials more closely mimic the non-stoichiometric nature of biological hydroxyapatite (HA). Typically, biological HA combines various CaP phases with different impurity ions, which substitute within the HA lattice, including strontium (Sr^{2+}), zinc (Zn^{2+}), magnesium (Mg^{2+}), carbonate (CO_3^{2-}) and fluoride (F^-), but to name a few. In addition to this biological HA have dimensions in the nanometre (nm) range, usually 60 nm in length by 5–20 nm wide. Both the effects of ion substitution and the nano-size crystals are seen as important factors for enhancing their potential biofunctionality. The driving hypothesis was to successfully synthesise nanoscale hydroxyapatite (nHA), co-substituted with strontium (Sr^{2+}) and zinc (Zn^{2+}) ions in varying concentrations using an aqueous precipitation method and to understand their chemical and physical properties. The materials were characterised using Fourier Transform Infrared Spectroscopy (FTIR), X-Ray Diffraction (XRD), X-Ray Photoelectron Spectroscopy (XPS) and Transmission Electron Microscopy (TEM) techniques. The FTIR, XRD and XPS results confirmed that the nHA was successfully co-substituted with Sr^{2+} and Zn^{2+} , replacing Ca^{2+} within the nHA lattice at varying concentrations. The FTIR results confirmed that all of the samples were carbonated, with a significant loss of hydroxylation as a consequence of the incorporation of Sr^{2+} and Zn^{2+} into the nHA lattice. The TEM results showed that each sample produced was nano-sized, with the Sr/Zn-10% nHA having the smallest sized crystals approximately 17.6 ± 3.3 nm long and 10.2 ± 1.4 nm wide. None of the materials synthesised here in this study contained any other impurity CaP phases. Therefore, this study has shown that co-substituted nHA can be prepared, and that the degree of substitution (and the substituting ion) can have a profound effect on the attendant materials' properties.

1. Introduction

Synthetic hydroxyapatite (HA) [$\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$] has been used extensively as a bone augmentation biomaterial, producing scaffolds for bone tissue engineering and for coating implants to promote bone healing and more effectively enhancing osseointegration [1,2]. It is well understood that nanoscale hydroxyapatite (nHA) with an average particle size of less than 20 nm can optimise the biofunctionality and bioactivity of HA, by increasing osteoblast cell proliferation [2].

Typically, biological apatite is non-stoichiometric and incorporates various impurity ions within the HA crystal lattice, Table 1 summarises the numerous substitutions which can occur, of those listed strontium and zinc have been shown to add significant enhancements to the properties of the HA individually. However, it has been suggested that co-substitutes of strontium and zinc could deliver a range of significant benefits over single ion substitutes [3,4]. Strontium and zinc co-

substituted within the nHA lattice can optimise osteoconductivity by assisting new bone growth and improve osteoinductivity, by encouraging the differentiation of mesenchymal stem cells to osteoblasts, which could work effectively to prevent osteoporosis [5–7]. Both ions have also been found to produce an antimicrobial response and zinc in particular may inhibit the growth of *Streptococcus mutans*, *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans* [8,9]. Therefore, both ions co-substituted together may also provide an effective antimicrobial treatment to combat conditions such as osteomyelitis [8].

The driving hypothesis of this work was to successfully synthesise nHA in a stoichiometric form, along with nHA co-substituted with Sr^{2+} and Zn^{2+} ions at a range of different weight percentage (wt%) concentrations, Sr/Zn-2.5% nHA, Sr/Zn-5% nHA and Sr/Zn-10% nHA. From previous research, one would expect the crystallinity, particle morphology and phase purity to vary as the level of ionic co-substitution was increased from 2.5% to 5–10% Sr/Zn [10,11]. Each sample was

* Correspondence to: Nanotechnology and Integrated Bioengineering Centre (NIBEC), School of Engineering, Ulster University, Room 22A08, Shore Road, Newtownabbey, Co. Antrim BT37 0QB, Northern Ireland, UK.

E-mail address: lowry-n2@ulster.ac.uk (N. Lowry).

<https://doi.org/10.1016/j.ceramint.2018.01.206>

Received 20 November 2017; Received in revised form 19 January 2018; Accepted 24 January 2018
0272-8842/ © 2018 Elsevier Ltd and Techna Group S.r.l. All rights reserved.

Table 1
Examples of various ions which can substitute within the HA lattice.

Substituted ion	Position of substitution	Ionic radius (Å) Ca (0.99Å)	Biological function and effect	Ref.
Strontium (Sr^{2+})	Ca^{2+}	1.12	High concentration where new bone is forming.	[5,6,8,9,32–34].
	Ca(I) Low Concentrations		Boosts osteoblast (OB) cell proliferation. Reduces osteoclast (OC) activity and therefore rate of bone resorption.	
Zinc (Zn^{2+})	Ca(II) High Concentrations	0.74	Encourages an antimicrobial effect	[6–8,30,31,35–39].
	Ca^{2+}		Inhibits OC resorption of bone. Encourages OB activity stimulating new bone formation. Strong antimicrobial effects.	
Magnesium (Mg^{2+})	Interstitial Insertion at Low Concentrations	0.72	Anti-inflammatory effects	[8,40–42].
	Ca(II) High Concentrations		Assists calcium ions to promote bone strength and health. Stimulates OB adhesion, proliferation and differentiation. Promotes osseointegration and bone ingrowth.	
Silver (Ag^+)	Ca^{2+}	1.26	Reduced β -TCP crystallinity, enhancing bone resorption	[8,43].
	Ca(I)		Exhibits an antibacterial effect on <i>Staphylococcus aureus</i> , <i>E-coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Candida albicans</i> and gram negative bacilli, with relatively low cytotoxicity. A minimum inhibitory concentration (MIC) of silver must be reached and maintained to inhibit later stage bacterial colonisation	
Lithium (Li^{+})	Ca^{2+}	0.76	Causes a reduction in bone remodelling, reducing bone turnover rates with lower amounts of bone loss.	[34,44–47].
			Stimulates the canonical Wnt signalling pathway, activating a major osteogenic signalling cascade, allowing MSCs to differentiate to OBs.	
Copper (Cu^{2+}) Silicon (SiO_4^{4-})	Ca^{2+}	0.73	Suppresses OC formation	[48,49]
	PO_4^{3-}		Antimicrobial effect against <i>E. coli</i> and <i>Candida albicans</i> .	
Carbonate (CO_3^{2-})	Substitutes for phosphate group at the 6 h position	5.86	Increases solubility and dissolution.	[8,30,50–52].
			Optimises cellular proliferation, neovascularisation and formation of mineral nodules and bone apposition. Encourages OB differentiation and collagen type-1 production in MG63 cells.	
Fluoride (F^-)	CO_3^{2-}	4.35	Osteoconductive silicon calcium phosphate scaffolds produce proliferation and differentiation of HBMSCs to matrix producing OBs, without any external osteogenic factors	[8,16,53].
	B type substitutes for phosphate ions (A/B ratio = 0.7–0.9)		Crystal morphology evolves as level of carbonate substitution increases from plate-like to needle-like to spherical.	
Fluoride (F^-)	OH^-	1.33	Reduced grain size and hardness value 200% larger than HA values, but not suitable for major load bearing applications.	[8,54]
	Substitutes for hydroxyl groups		Fast dissolution times, in vivo and in vitro due to carbonate weakening the bonds within the HA lattice. Bioactive and osseointegrative, forming a bone-like surface apatite layer in 7 days compared to 24–28 days for stoichiometric HA Reduces dental caries, dissolving in acidic environment of mouth after consumption of sugars and starches. Exhibits an antimicrobial effect. Boosts bone growth and formation, successfully treating osteoporosis.	

Table 2

Nomenclature used to describe samples for the study.

Sample name	Abbreviation used	Chemical formula	Molar quantity (mmol)				
			Ca(OH) ₂	H ₃ PO ₄	Ca(NO ₃) ₂ ·4H ₂ O	Sr(NO ₃) ₂ ·4H ₂ O	Zn(NO ₃) ₂ ·H ₂ O
Synthesised Nanoscale Hydroxyapatite	nHA	Ca ₁₀ (PO ₄) ₆ (OH) ₂	50.000	29.900	–	–	–
Synthesised Nanoscale Sr/Zn co- substituted Hydroxyapatite (1.25%/1.25%)	Sr/Zn-2.5%nHA	Ca _{9.75} Sr _{0.125} Zn _{0.125} (PO ₄) ₆ (OH) ₂	–	29.900	48.750	0.625	0.625
Synthesised Nanoscale Sr/Zn co- substituted Hydroxyapatite (2.5%/2.5%)	Sr/Zn-5%nHA	Ca _{9.5} Sr _{0.25} Zn _{0.25} (PO ₄) ₆ (OH) ₂	–	29.900	47.500	1.250	1.250
Synthesised Nanoscale Sr/Zn co- substituted Hydroxyapatite (5%/5%)	Sr/Zn-10%nHA	Ca ₉ Sr _{0.5} Zn _{0.5} (PO ₄) ₆ (OH) ₂	–	29.900	45.000	2.500	2.500

characterised and compared to the stoichiometric nHA using X-ray Diffraction (XRD), Fourier Transform Infrared Spectroscopy (FTIR), X-ray Photoelectron Spectroscopy (XPS) and Transmission Electron Microscopy (TEM). To the best of our knowledge, this paper represents one of the first attempts to produce, systematically characterise and compare the physicochemical properties of nano-hydroxyapatite, co-substituted to these various Sr²⁺/Zn²⁺ combined weight percentages, produced under identical conditions of pH and temperature.

2. Materials and methods

The nomenclature used to describe each of the calcium phosphate (CaP) powders synthesised in this study (using a standard aqueous precipitation method) are outlined in Table 2. Identical experimental conditions of temperature (60 °C) and pH ≥ 9, were used to synthesise pure nanoscale hydroxyapatite (nHA), nanoscale Sr(1.25%)/Zn(1.25%) hydroxyapatite (Sr/Zn-2.5%nHA), nanoscale Sr(2.5%)/Zn(2.5%) hydroxyapatite (Sr/Zn-5%nHA) and nanoscale Sr(5%)/Zn(5%) hydroxyapatite (Sr/Zn-10%nHA). Sodium hydroxide was used as a buffer to raise the pH ≥ 9 for the co-substitutions, Sr/Zn-2.5%nHA, Sr/Zn-5%nHA and Sr/Zn-10%nHA. The experimental ratios of Ca/P and (Ca + M)/P, (where M = Sr or Zn) were maintained at 1.67, for final comparison.

2.1. Aqueous precipitation synthesis of nHA

nHA powders were synthesised using an acid-base reaction of calcium hydroxide (Ca(OH)₂ – Sigma Aldrich) and phosphoric acid (85 wt % H₃PO₄ – Sigma Aldrich). Ca(OH)₂ was suspended in 500 ml of deionised H₂O, heated to 60 °C and stirred at 400 rpm for 1 h. An aqueous solution of H₃PO₄ was prepared by dissolving 85 wt% H₃PO₄ in 500 ml of deionised H₂O. The H₃PO₄ solution was added to the Ca(OH)₂ solution at a rate of 3.5 ml/min using a peristaltic pump. The solution was stirred for a further 2 h and allowed to settle and cool overnight. The pH of the solution was measured at 9.1. The precipitate was washed 3 times using deionised H₂O and then dried at 60 °C in an oven for 24 h.

2.2. Aqueous precipitation synthesis of strontium and zinc co-substituted nHA

Starting suspensions were prepared by dissolving calcium nitrate tetrahydrate (Ca(NO₃)₄·4H₂O – puriss.pa Sigma Aldrich) with both strontium nitrate (Sr(NO₃)₂·4H₂O – puriss pa. Sigma Aldrich) and zinc nitrate hexahydrate (Zn(NO₃)₂·H₂O – Sigma Aldrich) in 500 ml of deionised H₂O. The solution was stirred for 1 h at 60 °C and 400 rpm. An aqueous solution of H₃PO₄ was prepared by dissolving 85 wt% H₃PO₄ in 500 ml of deionised H₂O. The H₃PO₄ solution was added to the Ca(NO₃)₂·4H₂O solution at a rate of 3.5 ml/min using a peristaltic pump. The pH of each solution was typically measured between 2.0 and 2.5, with the solutions remaining colourless. On addition of the sodium

hydroxide to buffer each co-substituted solution to a pH ≥ 10, a white precipitate formed immediately. The solution was stirred for a further 2 h, with the pH maintained at ≥ 10 (by further addition of sodium hydroxide). The solution was allowed to settle and cool to room temperature overnight. The formed precipitate was washed 3 times using deionised H₂O and then dried at 60 °C in an oven for 24 h.

2.3. Characterisation of the nHA and co-substituted Sr/Zn-nHA powders

X-ray Diffraction (XRD) of the samples was carried out using a Bruker D8 Discover Diffractometer fitted with a Gobel mirror. Diffraction scans were obtained using a Cu Kα X-ray radiation (λ = 1.540 Å) source, with a tube voltage of 40 kV and tube current of 40 mA. Each diffraction scan was recorded at 2θ values from 5° to 70°, in 0.04° stepped increments, with a scan dwell time of 10 s for each increment.

Fourier Transform Infrared (FTIR) spectroscopy scans were obtained for each sample using a BIORAD FTS 3000MX Excalibur series instrument with a PIKE Diffuse Reflectance Infrared Fourier Transform Spectroscopy (DRIFTS) accessory. Samples were studied in absorbance mode from 4000 to 400 cm⁻¹, at a resolution of 4 cm⁻¹, with 20 scans per sample.

X-ray Photoelectron Spectroscopy (XPS) was performed using a Kratos Axis Ultra DLD Spectrometer. Spectra were analysed using monochromated Al Kα X-rays (hν = 1486.6 eV (eV)) operating at 10 kV and 15 mA (150 W). During analysis, a hybrid lens mode was used (electrostatic and magnetic) with a 300 μm × 700 μm analysis area. Wide energy survey scans (WESS) were taken at a pass energy of 160 eV, with high resolution spectra recorded at a pass energy of 20 eV. A Kratos charge neutraliser system with a filament current of 1.95 A and a charge balance between 3.3.3 V and 3.6 V, was used for all samples. Sample charging effects on the calculated BE positions were adjusted by setting the lowest BE component of the C1s spectral envelope to 285.0 eV, which is the generally accepted value for adventitious carbon surface contamination [12]. In total 3 areas were analysed upon each sample and a linear background was subtracted from each XPS spectra. The peak area of the most intense spectral lines for each elemental species was used to determine the percentage atomic concentration.

Transmission Electron Microscopy (TEM) observations were carried out using a JEOL 2100f microscope operated at 200 kV. Bright-field images were recorded from the powders supported on carbon films for size analysis. ImageJ software was utilised to measure the dimensions of 30 nanocrystals from each sample across 6 different TEM images. The results are reported as the mean ± standard deviation.

2.4. Statistical analysis

The statistical analysis for the TEM data was performed with GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego California USA. One way analysis of variance (ANOVA) was used

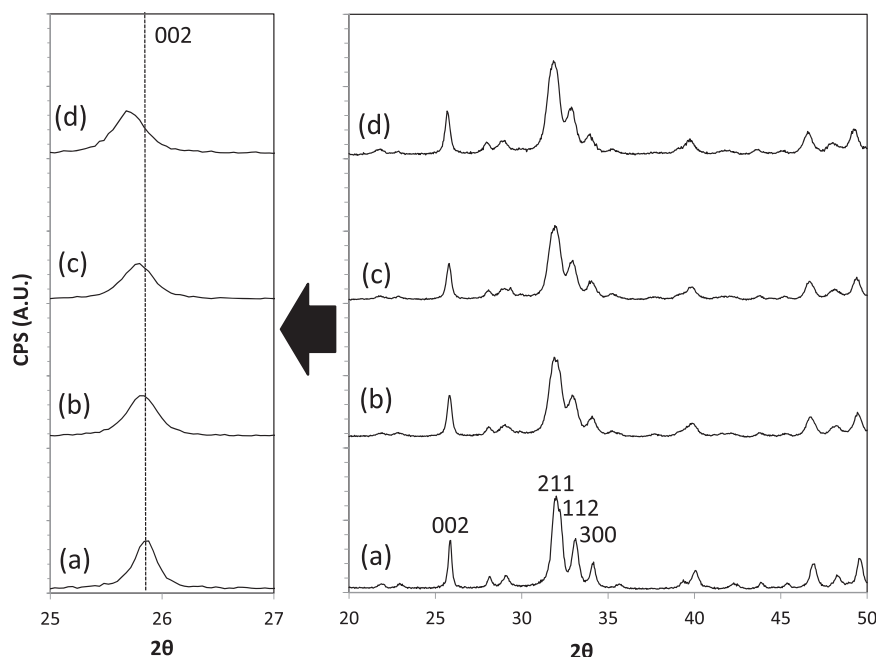


Fig. 1. XRD scans for nHA and co-substituted nHA samples.

to compare mean values in order to determine equivalence of variance between pairs of samples. Significance between groups was determined using the Bonferroni multiple comparison test. A value of $P < .05$ was taken as statistically significant.

3. Results

3.1. XRD

The comparative XRD patterns for the nHA, Sr/Zn2.5%nHA, Sr/Zn5%nHA and Sr/Zn10%nHA are shown in Fig. 1(a)–(d). The diffraction pattern for the nHA material is given in Fig. 1(a) and closely correlates to the International Centre for Diffraction Data (ICDD) file # 00-09-0432 for hydroxyapatite (HA). The peak positions and intensities recorded clearly indicate that the nHA material contains no detectable impurity phases. Fig. 1(b)–(d) for the Sr/Zn2.5%nHA, Sr/Zn5%nHA and Sr/Zn10%nHA samples, respectively, show diffraction peaks that are poorly resolved, indicating that each of the synthesised materials are nanoscale (≤ 100 nm), and/or have appreciable levels of substitutions within the nHA lattice, whereby Sr^{2+} and Zn^{2+} are substituting for Ca^{2+} . All 3 substituted nHA samples, Sr/Zn2.5%nHA, Sr/Zn5%nHA and Sr/Zn10%nHA exhibit noticeable shifts in XRD peak positions to lesser 2θ values, causing individual peak movement to the left, this is highlighted in Fig. 1 with movement of the 002 peak highlighted.

3.2. FTIR

FTIR spectra of the synthesised nHA and substituted nHA powders are shown in Fig. 2(a)–(d), respectively. Typical P–O stretching vibrations are clearly seen between 1200 and 900 cm^{-1} , while O–P–O bending vibrations can be observed between 610 and 400 cm^{-1} for each of the different materials [13]. O–H stretching and librational bands are also clearly visible around 3570 cm^{-1} and 632 cm^{-1} , respectively [14,15]. In addition to these vibrational bands, carbonate peaks (CO_3^{2-}) can also be observed between 1550 and 1400 cm^{-1} and around 870 cm^{-1} [16]. The carbonate peak positions may indicate that these different materials (both nHA and substituted nHA) have CO_3^{2-} ion substitutions at both the OH^- (A-site) and PO_4^{3-} (B-site) positions [13].

3.3. XPS

Figs. 3–6 show typical XPS wide energy survey scans (WESS) and high resolution scans recorded as B.E. (e.V.) versus intensity (counts), respectively, for the nHA, Sr/Zn-2.5%nHA, Sr/Zn-5%nHA and Sr/Zn10%nHA powders (namely the Ca2p, P2s, Sr3p and Zn2p peaks for each sample). Tables 3 and 4 quantify the XPS results recorded here. The main peaks recorded for the nHA sample were Ca2p_{3/2} (347.2 eV), P2p (133.7 eV), P2s (190.9 eV), and O1s (531.6 eV) are all clearly evident. The peak positions are as expected from previous reports, however, the Ca/P ratio was slightly lower than the expected 1.67 for stoichiometric HA, at 1.61 ± 0.04 as highlighted in Table 3 [13,15,17,18]. For all 3 substituted samples, the Ca/P ratios were calculated at 1.53 ± 0.06 for Sr/Zn-2.5%nHA, 1.42 ± 0.03 for Sr/Zn-5%nHA and 1.38 ± 0.01 for Sr/Zn-10%nHA, all of which are lower than 1.67 for stoichiometric HA. However, this is as expected and would be a consequence of the substitution of strontium and zinc for calcium within the nHA lattice [19–21].

As all 3 co-substituted samples contain strontium, it should be noted that the spectral envelope observed around 133.5 eV comprises an overlap of the Sr3d and P2p peaks as the Sr3d_{5/2} (133 ± 0.5 eV), Sr3d_{3/2} (135 ± 0.5 eV) and the P2p (132–133 eV) lines are positioned very close together. Therefore, to quantify the phosphorus and strontium atomic concentration % values in this study for all 3 co-substituted samples, the P2s and Sr3p peaks are used instead of the P2p and Sr3d peaks. The main peaks recorded for all 3 co-substituted samples, shown in Table 4 were therefore Ca2p_{3/2} (346.5–347.0 eV), P2s (190.0–190.3 eV), O1s (530.5–531.0 eV), Sr3p_{3/2} (268.3–268.8 eV), Zn2p_{3/2} (1021.7–1022.0 eV) and Na1s (1071.0–1071.5 eV). The Sr3p_{3/2} peaks confirm the presence and successful substitution of strontium for calcium within each co-substituted sample and the Zn2p_{3/2} peaks confirm the presence and successful substitution of zinc for calcium within each co-substituted sample.

3.4. TEM

The TEM images shown in Fig. 7 confirm that the nHA, Sr/Zn-2.5%nHA, Sr/Zn-5%nHA and Sr/Zn-10%nHA samples produced were all on the nanoscale, with the nHA crystals measuring 99.5 ± 12.9 nm in length and 33.8 ± 7.8 nm in breadth, with an aspect ratio of

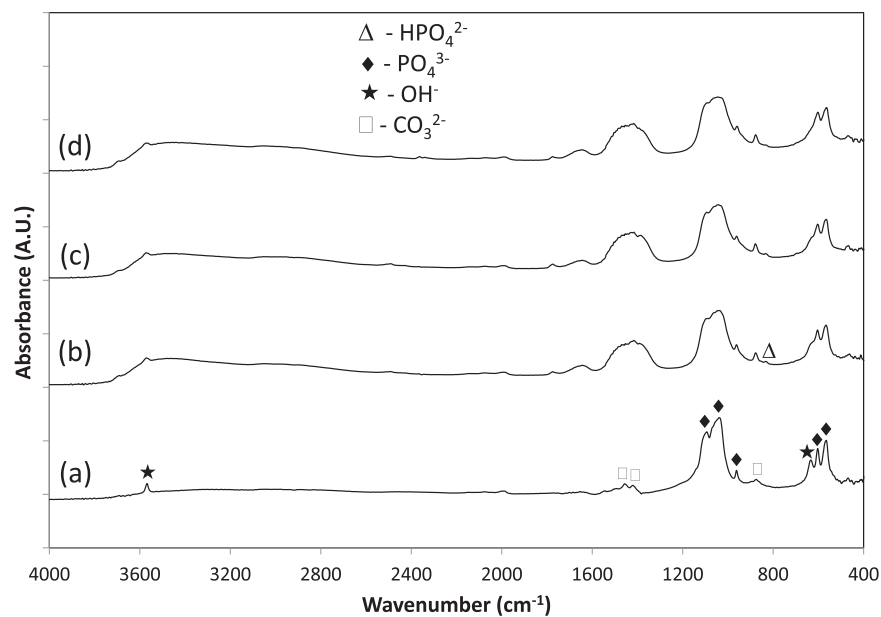


Fig. 2. FTIR spectra for nHA and co-substituted nHA samples.

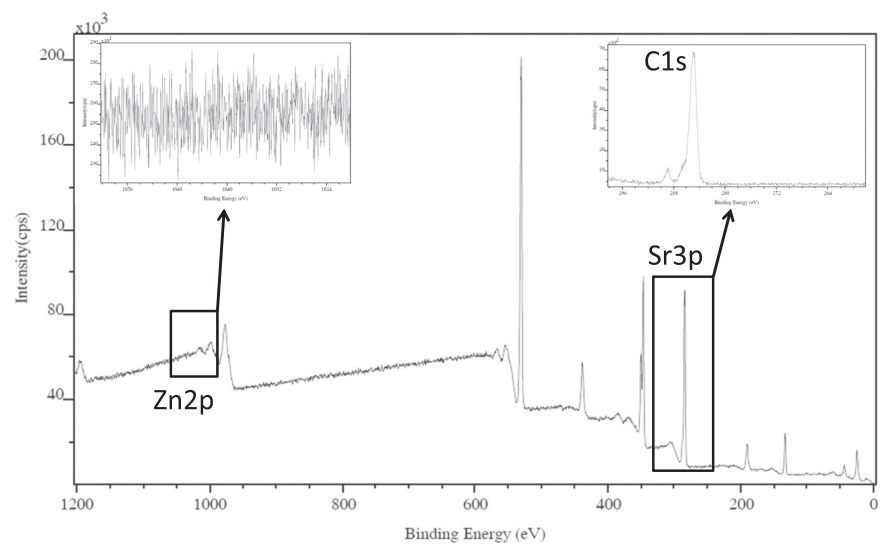


Fig. 3. XPS spectra for nHA sample.

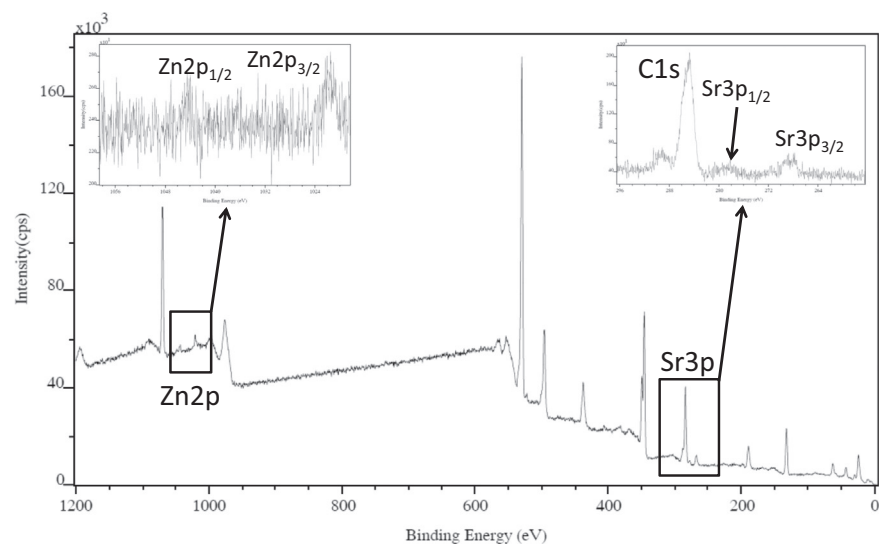


Fig. 4. XPS spectra for Sr/Zn2.5%nHA sample.

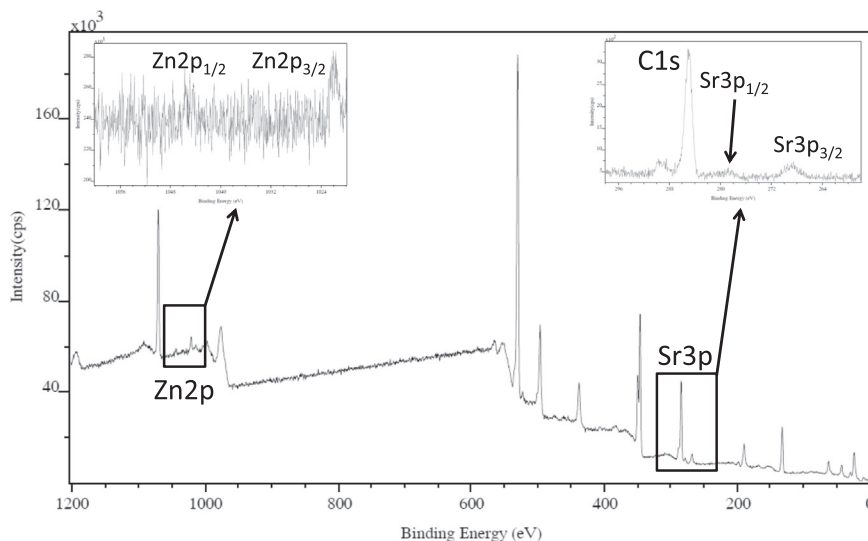


Fig. 5. XPS spectra for Sr/Zn5%nHA sample.

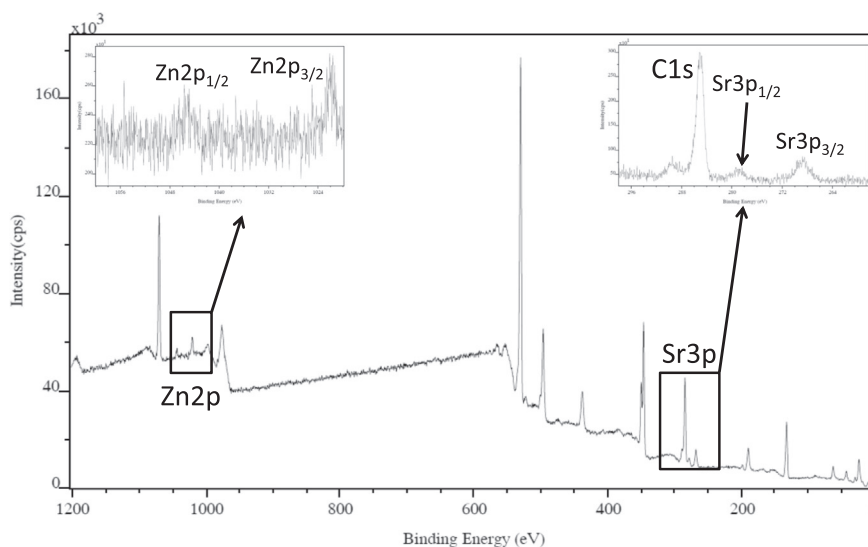


Fig. 6. XPS spectra for Sr/Zn10%nHA sample.

3.09 ± 0.75 , as reported in Table 5. The Sr/Zn-2.5%nHA, Sr/Zn-5%nHA and Sr/Zn-10%nHA samples all have nanocrystals, which are much smaller in size (both in length and breadth) than the nHA powder, as reported in Table 5, with typically lengths < 40 nm and breadths < 11 nm. The analysis of the TEM results, as highlighted in Fig. 8, show that there is a statistical difference in the length and breadth of the nHA, when compared with the co-substituted materials. In addition, within the co-substituted materials significant differences are observed in the length of the crystals between all samples. With respect to the aspect ratio, there was a significant difference between the nHA and the Sr/Zn-5%nHA and Sr/Zn-10%nHA samples (and between all of the different co-substituted materials). However, no significant statistical difference was observed in the aspect ratio between the nHA and the

Sr/Zn-2.5%nHA sample.

4. Discussion

4.1. XRD

The XRD diffraction patterns for all 3 co-substituted samples, Sr/Zn2.5%nHA, Sr/Zn5%nHA and Sr/Zn10%nHA in Fig. 1(b)–(d), show definite peak shifts to the left indicating that the diffraction peaks have shifted to slightly lower 2θ values, in comparison to the nHA sample. This is due to Sr^{2+} ions with a larger ionic radius (1.12Å) replacing Ca^{2+} ions with a smaller ionic radius (0.99Å). It can also be observed in the 002 peaks in Fig. 1 that as the wt% substitution of

Table 3
Comparative XPS ratios for standard nHA and substituted nHA powders (\pm standard deviation).

Sample	Ratios						
	Ca/P	(Ca + Sr)/P	(Ca + Zn)/P	(Ca + Sr + Zn)/P	Sr/(Sr + Ca)	Zn/(Zn + Ca)	Sr + Zn/(Sr + Zn + Ca)
nHA	1.61 ± 0.04	–	–	–	–	–	–
Sr/Zn-2.5%nHA	1.53 ± 0.06	1.59 ± 0.05	1.56 ± 0.06	1.63 ± 0.05	0.04 ± 0.01	0.02 ± 0.01	0.06 ± 0.01
Sr/Zn-5%nHA	1.42 ± 0.03	1.50 ± 0.04	1.47 ± 0.04	1.54 ± 0.05	0.05 ± 0.01	0.03 ± 0.01	0.08 ± 0.01
Sr/Zn-10%nHA	1.38 ± 0.01	1.55 ± 0.01	1.45 ± 0.04	1.63 ± 0.04	0.11 ± 0.01	0.05 ± 0.01	0.15 ± 0.02

Table 4

XPS peak positions for standard HA and substituted HA powders.

Sample	Peak positions – B.E. (eV)							
	C1s	O1s	Ca2p _{3/2}	P2p	P2s	Sr3p _{3/2}	Zn2p _{3/2}	Na1s
nHA	285.0	531.6	347.2	133.7	190.9	–	–	–
Sr/Zn-2.5%nHA	285.0	530.5	346.5	132.8	190.0	268.3	1021.7	1071.0
Sr/Zn-5%nHA	285.0	530.7	346.5	132.7	190.0	268.7	1021.7	1071.3
Sr/Zn10%nHA	285.0	531.0	347.0	133.0	190.3	268.8	1022.0	1071.5

strontium increases, from Fig. 1(b)–(d), so does the level of shifting to the left and lower 2 θ values increase, suggesting that gradually larger amounts of strontium have been successfully substituted into the nHA with each sample. The peaks also exhibited significant broadening with increasing Sr²⁺ content as can be seen in Fig. 1(b)–(d). Again, this result would be expected as increased Sr²⁺ substitution increases the strain within the lattice [3,22,23]. The XRD peak broadenings found in the 3 co-substituted samples may also indicate the successful substitution of Zn²⁺ ions for Ca²⁺ ions within the nHA lattice. However, peak positions would be expected to shift very slightly to higher 2 θ values, indicating the substitution of the smaller Zn²⁺ ion for the Ca²⁺ ion within the HA crystal lattice, causing a shrinkage in unit cell parameters. A peak shift to the right and higher 2 θ values cannot be observed in Fig. 1 but this could be due to the shift to the left and lower 2 θ values caused by the Sr²⁺ substitution within the same samples, masking any subsequent peak shift to the right caused by substituted Zn²⁺ ions. Previous research substituting HA with zinc alone has produced XRD diffraction patterns with these noticeable peak shifts to the right and higher 2 θ values, which could be used to confirm along with XPS data that Zn²⁺ ions have been successfully substituted within the nHA lattice however the larger amounts of Sr²⁺ ions substituted within the nHA lattice conceal any peak shift or increase in 2 θ value associated

with Zn²⁺ substitution [10,24,25].

4.2. FTIR

The FTIR peak positions observed for the nHA material for the P-O stretching and O-P-O bending vibrations were similar to those previously known for HA, as were those observed for the O-H librational and stretching modes. No peaks indicative of HPO₄²⁻ functional groups were detected for the nHA material (typically peaks for HPO₄²⁻ are seen around 1120 and 580 cm⁻¹) [17,19].

The 3 co-substituted samples appear to have broader P-O stretching bands located around 1099, 1043 and 964, and the O-P-O bending bands at 603 and 565, which could be due to both Sr²⁺ and Zn²⁺ substituting for calcium in the nHA lattices. There is also increased CO₃²⁻ substitution, in all 3 co-substituted samples, which previous literature has highlighted as an expected outcome when Zn²⁺ substitutes for Ca²⁺ in the nHA lattice [25,26]. In addition to these results, O-H stretching bands located around 3570 cm⁻¹ and O-H librational bands around 632 cm⁻¹ reduce in intensity for all 3 co-substituted samples in comparison to the nHA sample. This de-hydroxylation supports the fact that substitution of Sr²⁺ and Zn²⁺ ions for Ca²⁺ disrupts the nHA lattice.

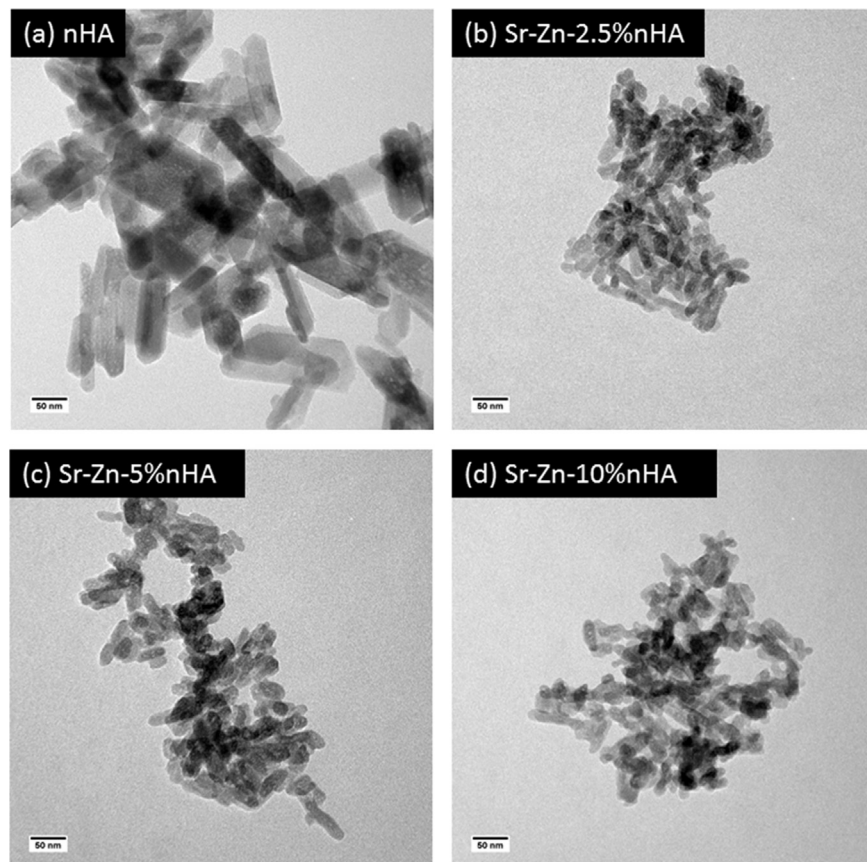


Fig. 7. TEM images of nHA and co-substituted nHA samples.

Table 5

Dimensions of nanocrystals (\pm standard deviation) measured using TEM analysis and processed using ImageJ software.

Sample	nHA	Sr/Zn-2.5%-nHA	Sr/Zn-5%-nHA	Sr/Zn-10%-nHA
Length (nm)	99.5 \pm 12.9	31.4 \pm 6.5	23.0 \pm 3.0	17.6 \pm 3.3
Breadth (nm)	33.8 \pm 7.8	9.5 \pm 1.5	10.0 \pm 1.5	10.2 \pm 1.4
Aspect ratio	3.09 \pm 0.75	3.40 \pm 0.96	2.36 \pm 0.47	1.75 \pm 0.33

4.3. XPS

The Ca/P ratio for the nHA sample was 1.61 ± 0.04 , slightly lower than the expected 1.67 for stoichiometric HA. All 3 co-substituted samples had further reduced Ca/P ratios, 1.53 ± 0.06 for Sr/Zn-2.5% nHA, 1.42 ± 0.03 for Sr/Zn-5% nHA and 1.38 ± 0.01 for Sr/Zn-10% nHA. This reduction in calcium content for each co-substituted sample would suggest successful substitution of strontium and zinc for calcium within the co-substituted nHA lattices, also as the wt% of strontium and zinc increases from 2.5%, 5% and 10%, the Ca/P ratio decreases, suggesting gradually more is substituted into the nHA, replacing calcium.

The (Ca + Sr)/P ratios for each co-substituted sample, found in Table 3 are as follows; 1.59 ± 0.05 for Sr/Zn-2.5% nHA, 1.50 ± 0.04 for Sr/Zn-5% nHA and 1.55 ± 0.01 for Sr/Zn-10% nHA. It can be seen that for each sample the ratio is larger for (Ca + Sr)/P in comparison to the Ca/P ratios alone, providing further evidence that for each co-substituted sample strontium has been successfully substituted for calcium within the nHA lattice. Again, the 3 (Ca + Zn)/P ratios for the co-substituted samples, Table 3 are as follows; 1.56 ± 0.06 for Sr/Zn-2.5% nHA, 1.47 ± 0.04 for Sr/Zn-5% nHA and 1.45 ± 0.04 for Sr/Zn-10% nHA. This would again confirm that zinc has successfully substituted for calcium within the nHA lattice as all 3 (Ca/Zn)/P ratios are larger than the previous Ca/P ratios for each corresponding co-substituted sample. This is again confirmed with the 3 co-substituted samples (Ca + Sr + Zn)/P ratios, found in Table 3 which are; 1.63 ± 0.05 for Sr/Zn-2.5% nHA, 1.54 ± 0.05 for Sr/Zn-5% nHA and 1.63 ± 0.04 for Sr/Zn-10% nHA, which are again larger figures than the previous Ca/P, (Ca + Sr)/P and (Ca + Zn)/P ratios for each individual sample, again confirming the successful substitution of both strontium and zinc for calcium within each nHA lattice.

The Sr/(Sr + Ca) ratios found in Table 3 are interesting as they show a clear increase in strontium substitution for each increasing wt% sample, therefore, for Sr/Zn-2.5% nHA, 0.04 ± 0.01 , for Sr/Zn-5% nHA, 0.05 ± 0.01 and for Sr/Zn-10% nHA, 0.11 ± 0.01 . This data indicates that as the wt% concentration of strontium is increased from 1.25% to

2.5% to 5% for each co-substituted sample the subsequent percentage ratios of atomic concentration also increase accordingly, confirming that as a larger percentage concentration of strontium is added at the beginning of the reaction, a larger amount of strontium is substituted for calcium within the nHA lattice of each sample. A similar trend can be seen with the Zn/(Zn + Ca) ratios again located in Table 3, a clear increase in zinc substitution for each increasing wt% sample can be seen, for Sr/Zn-2.5% nHA, 0.02 ± 0.01 , for Sr/Zn-5% nHA, 0.03 ± 0.01 and for Sr/Zn-10% nHA, 0.05 ± 0.01 . This would again indicate that as the wt% concentration of zinc is increased from 1.25%, 2.5% to 5% for each co-substituted sample the subsequent percentage ratios of atomic concentration also increase, again confirming that as a larger percentage concentration of zinc is added at the beginning of each synthesis, a larger amount of zinc is substituted for calcium within the nHA lattice of each sample. Again, the Sr + Zn/(Sr + Zn + Ca) ratios for Sr/Zn-2.5% nHA, 0.06 ± 0.01 , for Sr/Zn-5% nHA, 0.08 ± 0.01 and for Sr/Zn-10% nHA, 0.15 ± 0.02 agree with the strontium and zinc ratios and show a distinct rise in % atomic concentration, as the starting percentage concentrations of both strontium and zinc are increased. This indicates that a larger amount of both strontium and zinc has been co-substituted into the nHA lattice of each sample, as the starting concentrations of strontium and zinc increase from 1.25% to 2.5% to 5% for each ion. This suggests that the wet precipitation synthesis successfully substitutes larger concentrations of both strontium and zinc within the nHA lattice as amounts of the starting strontium nitrate and zinc nitrate hexahydrate are increased.

Small amounts of sodium were also detected in the XPS spectra analyses, with the Na1s peak located between 1071.0 and 1071.5 eV for all 3 co-substituted samples. This was caused by the use of sodium hydroxide as a buffer to increase the pH above 10 during each precipitation reaction and has been found to encourage the substitution of zinc for calcium within the nHA lattice. It should be noted that XPS is a surface analytical technique and only penetrates the uppermost surface (1–10 μ m) of each sample.

4.4. TEM

The TEM results show clearly that as the co-substituted wt% of strontium and zinc increased from 2.5, 5 to 10%, the average particle length for each sample reduced from 31.4 ± 6.5 nm, 23.0 ± 3.0 nm and 17.6 ± 3.3 nm, respectively with the breadth remaining relatively constant at 9.5 ± 1.5 nm, 10.0 ± 1.5 nm and 10.2 ± 1.4 nm, respectively. This corresponds with previous research which showed that nHA substituted with both strontium and zinc produced smaller sized particle samples [27,28]. It has also been reported in previous literature that as the concentration of the substituted ion rises, regardless of

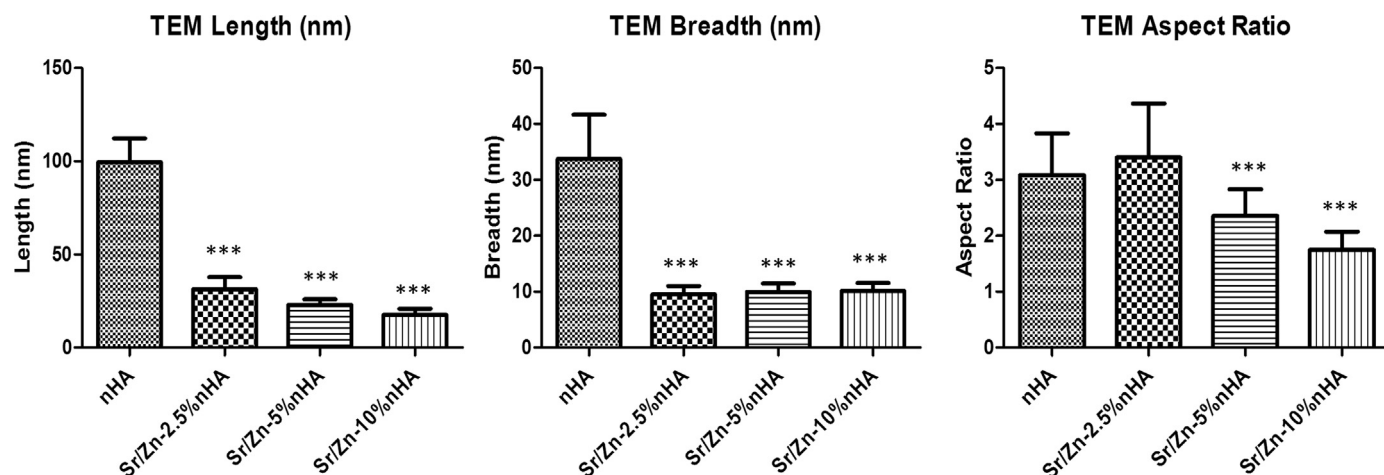


Fig. 8. TEM statistical comparisons between nHA and co-substituted Sr/Zn nHA samples. *** Statistically significant at $P < .0001$ in comparison with the control nHA.

whether it is strontium or zinc, the subsequent substituted nHA crystals tend to agglomerate [27,28]. The statistical analysis of the crystal length, breadth and aspect ratio here, highlight that as the concentration of the co-substituted ions increase, it has an obvious effect on the growth of the crystals, (with higher ion substitutions inhibiting crystal growth).

This study has also found that for every sample analysed using TEM, the observed nanocrystals appeared granular in nature, which may indicate a high level of porosity and an increased surface area within both the nHA and substituted nHA materials synthesised. An optimised surface nano-topography could in turn increase adsorption of specific proteins and ultimately improve osteoblast cell adhesion and promote osteoblast proliferation and differentiation [29]. Previous research has also found that nano-sized HA, with a larger surface area, has increased sinterability and densification which can in turn improve its mechanical properties and fracture toughness [2,29].

5. Conclusion

The specific aim of this work was to synthesise nHA and nHA co-substituted with strontium and zinc at varying concentrations, Sr/Zn-2.5%nHA, Sr/Zn-5%nHA and Sr/Zn-10%nHA, using an aqueous precipitation technique. The results indicated that nHA and nHA co-substituted with strontium and zinc were successfully produced at varying concentrations. The nHA materials produced did not contain any other CaP impurity phases and were proven to be nano-sized in scale, as shown by XRD, FTIR, XPS and TEM analyses. Further comparison did indicate that crystallinity did vary as the amount of Sr^{2+} and Zn^{2+} increased within the co-substituted samples, with the degree of structural order decreasing as Ca^{2+} ions were progressively replaced with either larger Sr^{2+} or smaller Zn^{2+} ions with increasing dehydroxylation observed within the co-substituted samples, which further highlighted disruption of the HA lattice. Also, the particle morphology of the co-substituted nHA samples changed with increased co-substitution, with the particles steadily reducing in length.

Very few previous studies have investigated the synthesis of nHA materials co-substituted with Sr^{2+} and Zn^{2+} . Therefore, the successful co-substitution of both strontium and zinc within nHA, as demonstrated successfully here, could provide a more bioactive bone substitute biomaterial. However, there is a need for further detailed in vitro and in vivo studies to confirm the efficacy of these co-substituted materials.

Acknowledgements

The authors would like to acknowledge the Ph.D. funding provided by the Department for the Economy (Northern Ireland) to support this work.

References

- [1] K. Fox, P.A. Tran, N. Tran, Recent advances in research applications of nanophase hydroxyapatite, *ChemPhysChem* 13 (2012) 2495–2506, <http://dx.doi.org/10.1002/cphc.201200080>.
- [2] H. Zhou, J. Lee, Nanoscale hydroxyapatite particles for bone tissue engineering, *Acta Biomater.* 7 (2011) 2769–2781, <http://dx.doi.org/10.1016/j.actbio.2011.03.019>.
- [3] A. Siddharthan, S.K. Seshadri, T.S.S. Kumar, Rapid synthesis of calcium deficient hydroxyapatite nanoparticles by microwave irradiation, *Trends Biomater. Artif. Organs* 18 (2005) 110–113, <http://dx.doi.org/10.1016/j.ssc.2004.02.045>.
- [4] I.R. de Lima, G.G. Alves, C.A. Soriano, A.P. Campanelli, T.H. Gasparoto, E. Schneider Ramos, L.A. de Sena, A.M. Rossi, J.M. Granjeiro, Understanding the impact of divalent cation substitution on hydroxyapatite: an in vitro multiparametric study on biocompatibility, *J. Biomed. Mater. Res. Part A* 98A (2011) 351–358, <http://dx.doi.org/10.1002/jbm.a.33126>.
- [5] E. Bonnelye, A. Chabadel, F. Saltel, P. Jurdic, Dual effect of strontium ranelate: stimulation of osteoblast differentiation and inhibition of osteoclast formation and resorption in vitro, *Bone* 42 (2008) 129–138, <http://dx.doi.org/10.1016/j.bone.2007.08.043>.
- [6] M. Roy, G.A. Fielding, A. Bandyopadhyay, S. Bose, Effects of zinc and strontium substitution in tricalcium phosphate on osteoclast differentiation and resorption, *Biomater. Sci.* (2013) 74–82, <http://dx.doi.org/10.1039/c2bm00012a>.
- [7] D.V. Shepherd, K. Kauppinen, R.A. Brooks, S.M. Best, An in vitro study into the effect of zinc substituted hydroxyapatite on osteoclast number and activity, *J. Biomed. Mater. Res. – Part A* 102 (2014) 4136–4141, <http://dx.doi.org/10.1002/jbm.a.35089>.
- [8] J.H. Shepherd, D.V. Shepherd, S.M. Best, Substituted hydroxyapatites for bone repair, *J. Mater. Sci. Mater. Med.* 23 (2012) 2335–2347, <http://dx.doi.org/10.1007/s10856-012-4598-2>.
- [9] V. Aina, L. Bergandi, G. Lusvardi, G. Malavasi, F.E. Imrie, I.R. Gibson, G. Cerrato, D. Ghigo, Sr-containing hydroxyapatite: morphologies of HA crystals and bioactivity on osteoblast cells, *Mater. Sci. Eng. C* 33 (2013) 1132–1142, <http://dx.doi.org/10.1016/j.msec.2012.12.005>.
- [10] N. Lowry, Y. Han, B.J. Meenan, A.R. Boyd, Strontium and zinc co-substituted nanophase hydroxyapatite, *Ceram. Int.* 43 (2017) 12070–12078, <http://dx.doi.org/10.1016/j.ceramint.2017.06.062>.
- [11] L. Robinson, K. Salma-Ancane, L. Stipniece, B.J. Meenan, A.R. Boyd, The deposition of strontium and zinc Co-substituted hydroxyapatite coatings, *J. Mater. Sci. Mater. Med.* 28 (2017), <http://dx.doi.org/10.1007/s10856-017-5846-2>.
- [12] L. Tortet, J.R. Gavarri, G. Nihoul, Study of Protonic Mobility in $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$ (Brushite) and CaHPO_4 (Monelite) By Infrared Spectroscopy and Neutron Scattering, 16, 1997, pp. 6–16.
- [13] A.R. Boyd, C. O’Kane, B.J. Meenan, Control of calcium phosphate thin film stoichiometry using multi-target sputter deposition, *Surf. Coat. Technol.* 233 (2013) 131–139, <http://dx.doi.org/10.1016/j.surfcoat.2013.04.017>.
- [14] C.-J. Chung, H.-Y. Long, Systematic strontium substitution in hydroxyapatite coatings on titanium via micro-arc treatment and their osteoblast/osteoclast responses, *Acta Biomater.* 7 (2011) 4081–4087, <http://dx.doi.org/10.1016/j.actbio.2011.07.004>.
- [15] A.R. Boyd, B.J. Meenan, N.S. Leyland, Surface characterisation of the evolving nature of radio frequency (RF) magnetron sputter deposited calcium phosphate thin films after exposure to physiological solution, *Surf. Coat. Technol.* 200 (2006) 6002–6013, <http://dx.doi.org/10.1016/j.surfcoat.2005.09.032>.
- [16] I.R. Gibson, W. Bonfield, Novel synthesis and characterization of an AB-type carbonate-substituted hydroxyapatite, *J. Biomed. Mater. Res.* 59 (2002) 697–708, <http://dx.doi.org/10.1002/jbm.10044>.
- [17] A.R. Boyd, L. Rutledge, L.D. Randolph, B.J. Meenan, Strontium-substituted hydroxyapatite coatings deposited via a co-deposition sputter technique, *Mater. Sci. Eng. C* 46 (2015) 290–300, <http://dx.doi.org/10.1016/j.msec.2014.10.046>.
- [18] A. Costescu, I. Pasuk, F. Ungureanu, A. Dinischiotu, F. Huneau, S. Galaup, P.L.E. Coustumer, D. Predoi, C. Ftir, Physico-Chemical Properties of Nano-Sized Hexagonal Hydroxyapatite Powder Synthesized by Sol-Gel, 5, 2010, pp. 89–1000.
- [19] A.R. Boyd, L. Rutledge, L.D. Randolph, I. Mutreja, B.J. Meenan, The deposition of strontium-substituted hydroxyapatite coatings, *J. Mater. Sci. Mater. Med.* 26 (2015) 65, <http://dx.doi.org/10.1007/s10856-014-5377-z>.
- [20] W. Xia, C. Lindahl, C. Persson, P. Thomsen, J. Lausmaa, H. Engqvist, Changes of Surface Composition and Morphology after Incorporation of Ions into Biomimetic Apatite Coatings, 2010, 2010, pp. 7–16. <https://dx.doi.org/10.4236/jbmb.2010.11002>.
- [21] Y. Zhao, D. Guo, S. Hou, H. Zhong, J. Yan, C. Zhang, Y. Zhou, Porous Allograft Bone Scaffolds: Doping with Strontium, 8, 2013, pp. 1–10. <https://dx.doi.org/10.1371/journal.pone.0069339>.
- [22] Y.Y. Özbek, F.E. Baştan, F. Üstel, Synthesis and characterization of strontium-doped hydroxyapatite for biomedical applications, *J. Therm. Anal. Calorim.* 125 (2016) 745–750, <http://dx.doi.org/10.1007/s10973-016-5607-3>.
- [23] L. Li, X. Lu, Y. Meng, C.M. Weyant, Comparison study of biomimetic strontium-doped calcium phosphate coatings by electrochemical deposition and air plasma spray: morphology, composition and bioactive performance, *J. Mater. Sci. Mater. Med.* 23 (2012) 2359–2368, <http://dx.doi.org/10.1007/s10856-012-4633-3>.
- [24] A. Anwar, S. Akbar, A. Sadiqa, M. Kazmi, Novel continuous flow synthesis, characterization and antibacterial studies of nanoscale zinc substituted hydroxyapatite bioceramics, *Inorg. Chim. Acta* 453 (2016) 16–22, <http://dx.doi.org/10.1016/j.ica.2016.07.041>.
- [25] I. Pereiro, C. Rodriguez-Valencia, C. Serr, C. Solla, J. Serra, P. Gonzalez, Structural properties of ZnO films grown by picosecond pulsed-laser deposition, *Appl. Surf. Sci.* 258 (2012) 9192–9197.
- [26] M. Kavitha, R. Subramanian, R. Narayanan, V. Udhayabanu, Solution combustion synthesis and characterization of strontium substituted hydroxyapatite nanocrystals, *Powder Technol.* 253 (2014) 129–137, <http://dx.doi.org/10.1016/j.powtec.2013.10.045>.
- [27] V. Krishnan, A. Bhatia, H. Varma, Development, characterization and comparison of two strontium doped nano hydroxyapatite molecules for enamel repair / regeneration, *Dent. Mater.* 32 (2016) 646–659, <http://dx.doi.org/10.1016/j.dental.2016.02.002>.
- [28] K.P. Tank, P. Sharma, D.K. Kanchan, M.J. Joshi, FTIR, Powder XRD, TEM and Dielectric Studies of Pure and Zinc Doped Nano-Hydroxyapatite, 1316, 2011, pp. 1309–1316. <https://dx.doi.org/10.1002/crat.201100080>.
- [29] S.V. Dorozhkin, Nanosized and nanocrystalline calcium orthophosphates, *Acta Biomater.* 6 (2010) 715–734, <http://dx.doi.org/10.1016/j.actbio.2009.10.031>.
- [30] H. Zreiqat, Y. Ramaswamy, C. Wu, A. Paschalidis, Z. Lu, B. James, O. Birke, M. McDonald, D. Little, C.R. Dunstan, The incorporation of strontium and zinc into a calcium–silicon ceramic for bone tissue engineering, *Biomaterials* 31 (2010) 3175–3184, <http://dx.doi.org/10.1016/j.biomaterials.2010.01.024>.
- [31] V. Mourino, J.P. Cattalini, A.R. Boccacini, Metallic ions as therapeutic agents in tissue engineering scaffolds: an overview of their biological applications and strategies for new developments, *J. R. Soc. Interface* 9 (2012) 401–419, <http://dx.doi.org/10.1098/rsif.2011.0611>.

- [32] F. Yang, D. Yang, J. Tu, Q. Zheng, L. Cai, L. Wang, Strontium enhances osteogenic differentiation of mesenchymal stem cells and in vivo bone formation by activating Wnt/catenin signaling, *Stem Cells* (2011), <http://dx.doi.org/10.1002/stem.646>.
- [33] T. Kubota, T. Michigami, K. Ozono, Wnt signaling in bone metabolism, *J. Bone Miner. Metab.* 27 (2009) 265–271, <http://dx.doi.org/10.1007/s00774-009-0064-8>.
- [34] M. Arioka, F. Takahashi-Yanaga, M. Sasaki, T. Yoshihara, S. Morimoto, M. Hirata, Y. Mori, T. Sasaguri, Acceleration of bone regeneration by local application of lithium: Wnt signal-mediated osteoblastogenesis and Wnt signal-independent suppression of osteoclastogenesis, *Biochem. Pharmacol.* 90 (2014) 397–405, <http://dx.doi.org/10.1016/j.bcp.2014.06.011>.
- [35] S.C. Cox, P. Jamshidi, L.M. Grover, K.K. Mallick, Preparation and characterisation of nanophase Sr, Mg, and Zn substituted hydroxyapatite by aqueous precipitation, *Mater. Sci. Eng. C* 35 (2014) 106–114, <http://dx.doi.org/10.1016/j.msec.2013.10.015>.
- [36] F. Ren, R. Xin, X. Ge, Y. Leng, Characterization and structural analysis of zinc-substituted hydroxyapatites, *Acta Biomater.* 5 (2009) 3141–3149, <http://dx.doi.org/10.1016/j.actbio.2009.04.014>.
- [37] H. Storrie, S.I. Stupp, Cellular Response to Zinc-Containing Organoapatite: An In Vitro Study of Proliferation, Alkaline Phosphatase Activity and Biomineralization, 26, 2005, pp. 5492–5499. <https://dx.doi.org/10.1016/j.biomaterials.2005.01.043>.
- [38] G.S. Kumar, A. Thamizhavel, Y. Yokogawa, S.N. Kalkura, E.K. Giriya, Synthesis, characterization and in vitro studies of zinc and carbonate co-substituted nano-hydroxyapatite for biomedical applications, *Mater. Chem. Phys.* 134 (2012) 1127–1135, <http://dx.doi.org/10.1016/j.matchemphys.2012.04.005>.
- [39] R.J. Friederichs, H.F. Chappell, D.V. Shepherd, S.M. Best, Synthesis, characterization and modelling of zinc and silicate co-substituted hydroxyapatite, *J. R. Soc. Interface* 12 (2015) 20150190, <http://dx.doi.org/10.1098/rsif.2015.0190>.
- [40] L. Stipniece, K. Salma-Ancane, N. Borodajenko, M. Sokolova, D. Jakovlevs, L. Berzina-Cimdina, Characterization of Mg-substituted hydroxyapatite synthesized by wet chemical method, *Ceram. Int.* 40 (2014) 3261–3267, <http://dx.doi.org/10.1016/j.ceramint.2013.09.110>.
- [41] M. Vandrovcova, T.E.L. Douglas, W. Mróz, O. Musial, D. Schaubroeck, B. Budner, R. Syroka, P. Dubrue, L. Bacakova, Pulsed laser deposition of magnesium-doped calcium phosphate coatings on porous polycaprolactone scaffolds produced by rapid prototyping, *Mater. Lett.* 148 (2015) 178–183, <http://dx.doi.org/10.1016/j.matlet.2015.02.074>.
- [42] K. Salma-Ancane, L. Stipniece, A. Putnins, L. Berzina-Cimdina, Development of Mg-containing porous β -tricalcium phosphate scaffolds for bone repair, *Ceram. Int.* 41 (2015) 4996–5004, <http://dx.doi.org/10.1016/j.ceramint.2014.12.065>.
- [43] G.A. Fielding, M. Roy, A. Bandyopadhyay, S. Bose, Antibacterial and biological characteristics of silver containing and strontium doped plasma sprayed hydroxyapatite coatings, *Acta Biomater.* 8 (2012) 3144–3152, <http://dx.doi.org/10.1016/j.actbio.2012.04.004>.
- [44] A. Zamani, G.R. Omrani, M.M. Nasab, Lithium's effect on bone mineral density, *Bone* 44 (2009) 331–334, <http://dx.doi.org/10.1016/j.bone.2008.10.001>.
- [45] H.D. Jang, J.H. Shin, D.R. Park, J.H. Hong, K. Yoon, R. Ko, C.Y. Ko, H.S. Kim, D. Jeong, N. Kim, S.Y. Lee, Inactivation of glycogen synthase kinase-3 β is required for osteoclast differentiation, *J. Biol. Chem.* 286 (2011) 39043–39050, <http://dx.doi.org/10.1074/jbc.M111.256768>.
- [46] J. Albers, J. Keller, A. Baranowsky, F.T. Beil, P. Catala-Lehnen, J. Schulze, M. Amling, T. Schinke, Canonical Wnt signaling inhibits osteoclastogenesis independent of osteoprotegerin, *J. Cell Biol.* 200 (2013) 537–549, <http://dx.doi.org/10.1083/jcb.201207142>.
- [47] W. Wei, D. Zeve, J.M. Suh, X. Wang, Y. Du, J.E. Zerwekh, P.C. Dechow, J.M. Graff, Y. Wan, Biphasic and dosage-dependent regulation of osteoclastogenesis by -catenin, *Mol. Cell. Biol.* 31 (2011) 4706–4719, <http://dx.doi.org/10.1128/MCB.05980-11>.
- [48] V. Stanic, S. Dimitrijevic, J. Antic-Stankovic, M. Mitric, B. Jokic, I.B. Plecac, S. Raicevic, Synthesis, characterization and antimicrobial activity of copper and zinc-doped hydroxyapatite nanopowders, *Appl. Surf. Sci.* 256 (2010) 6083–6089, <http://dx.doi.org/10.1016/j.apsusc.2010.03.124>.
- [49] W.-L. Du, Y.-L. Xu, Z.-R. Xu, C.-L. Fan, Preparation, characterization and antibacterial properties against *E. coli* K(88) of chitosan nanoparticle loaded copper ions, *Nanotechnology* 19 (2008) 85707, <http://dx.doi.org/10.1088/0957-4484/19/8/085707>.
- [50] R.J. Friederichs, R.A. Brooks, M. Ueda, S.M. Best, In vitro osteoclast formation and resorption of silicon-substituted hydroxyapatite ceramics, *J. Biomed. Mater. Res. – Part A* 103 (2015) 3312–3322, <http://dx.doi.org/10.1002/jbm.a.35470>.
- [51] D.M. Reffitt, N. Ogston, R. Jugdaohsingh, H.F.J. Cheung, B.A.J. Evans, R.P.H. Thompson, J.J. Powell, G.N. Hampson, Orthosilicic Acid Stimulates Collagen Type 1 Synthesis and Osteoblastic Differentiation in Human Osteoblast-Like Cells In Vitro, 32, 2003, pp. 127–135. [https://dx.doi.org/10.1016/S8756-3282\(02\)00950-X](https://dx.doi.org/10.1016/S8756-3282(02)00950-X).
- [52] R. Ferro De Godoy, S. Hutchens, C. Campion, G. Blunn, Silicate-substituted calcium phosphate with enhanced strut porosity stimulates osteogenic differentiation of human mesenchymal stem cells, *J. Mater. Sci. Mater. Med.* 26 (2015) 5387, <http://dx.doi.org/10.1007/s10856-015-5387-5>.
- [53] J.C. Merry, I.R. Gibson, S.M. Best, W. Bonfield, Synthesis and characterization of carbonate hydroxyapatite, *J. Mater. Sci. Mater. Med.* 9 (1998) 779–783, <http://dx.doi.org/10.1023/A:1008975507498>.
- [54] V. Stanic, S. Dimitrijevic, D.G. Antonovic, B.M. Jokic, S.P. Zec, S.T. Tanaskovic, S. Raicevic, Synthesis of fluorine substituted hydroxyapatite nanopowders and application of the central composite design for determination of its antimicrobial effects, *Appl. Surf. Sci.* 290 (2014) 346–352, <http://dx.doi.org/10.1016/j.apsusc.2013.11.081>.